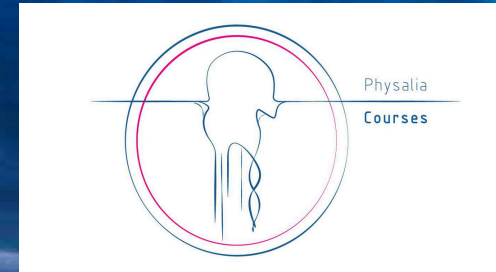


Introduction to Trinity RNA-Seq



Berlin, June 2017

Brian Haas
Broad Institute



Welcome to the Berlin 2017 Trinity Workshop Wiki!

Day	Time	Activities
Monday, June 12	morning	Workshop Introduction
		Exploring the Computational Infrastructure
	afternoon	Unix command-line review
		Data overview and setup
		Using FASTQC and Trimmomatic
Tuesday, June 13	morning	Trinity de novo transcriptome assembly
		Expression quantification
	afternoon	Quality assessment for assembly
		QC samples and replicates
Wednesday, June 14	morning	Statistical methods for differential expression analysis
	afternoon	Transcript clustering and expression profiling
Thursday, June 15	morning	Methods for functional annotation
	afternoon	Trinotate and TrinotateWeb
	afternoon	Functional enrichment analysis
Friday, June 16	morning	Review and custom data analyses

<https://github.com/trinityrnseq/BerlinTrinityWorkshop2017/wiki>

Generating RNA-Seq: *How to Choose?*

Many different instruments hit the scene in the last decade



Illumina



454



SOLiD



Helicos



Ion Torrent

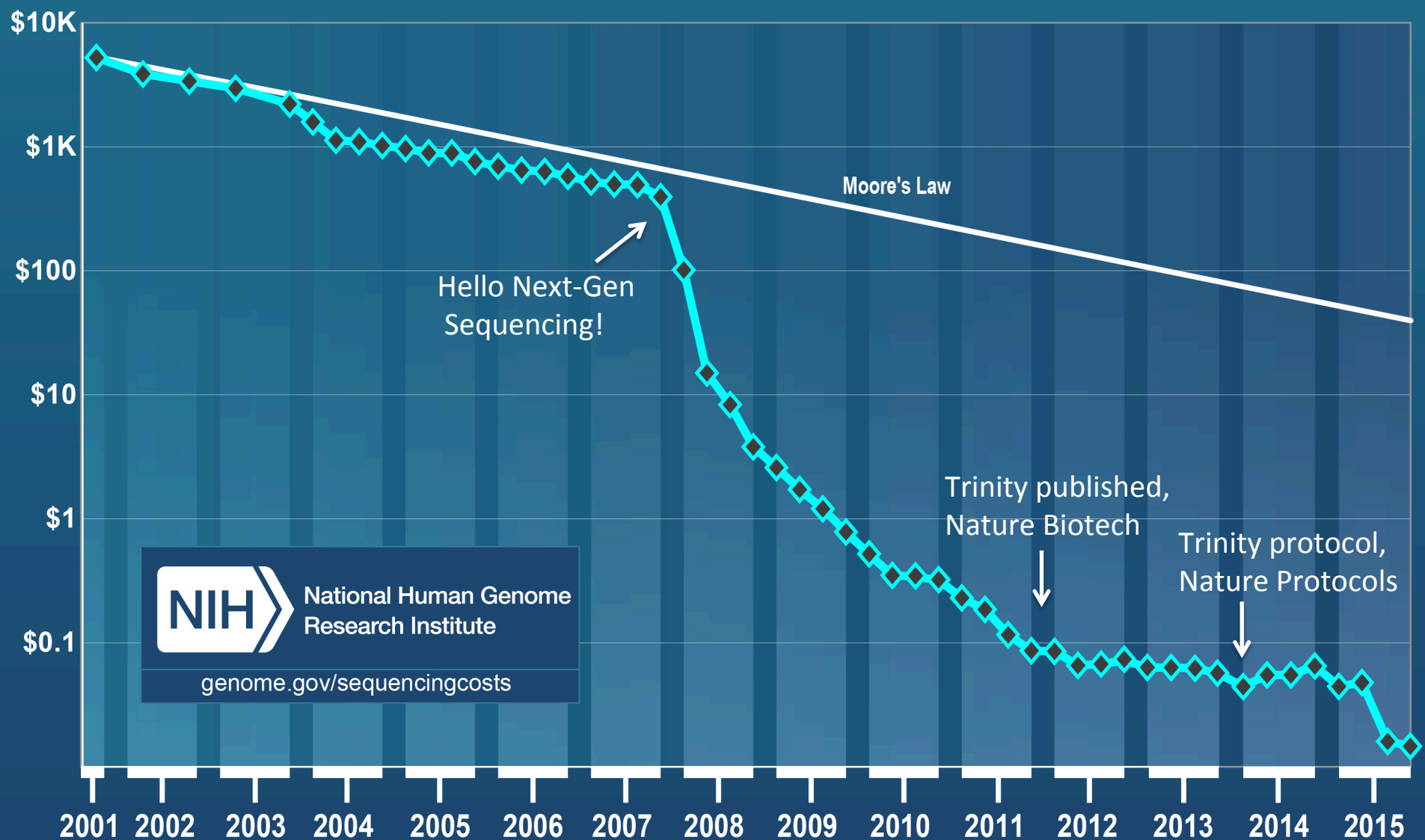


Pacific Biosciences



Oxford Nanopore

Cost per Raw Megabase of DNA Sequence



From <https://www.genome.gov/sequencingcostsdata/>

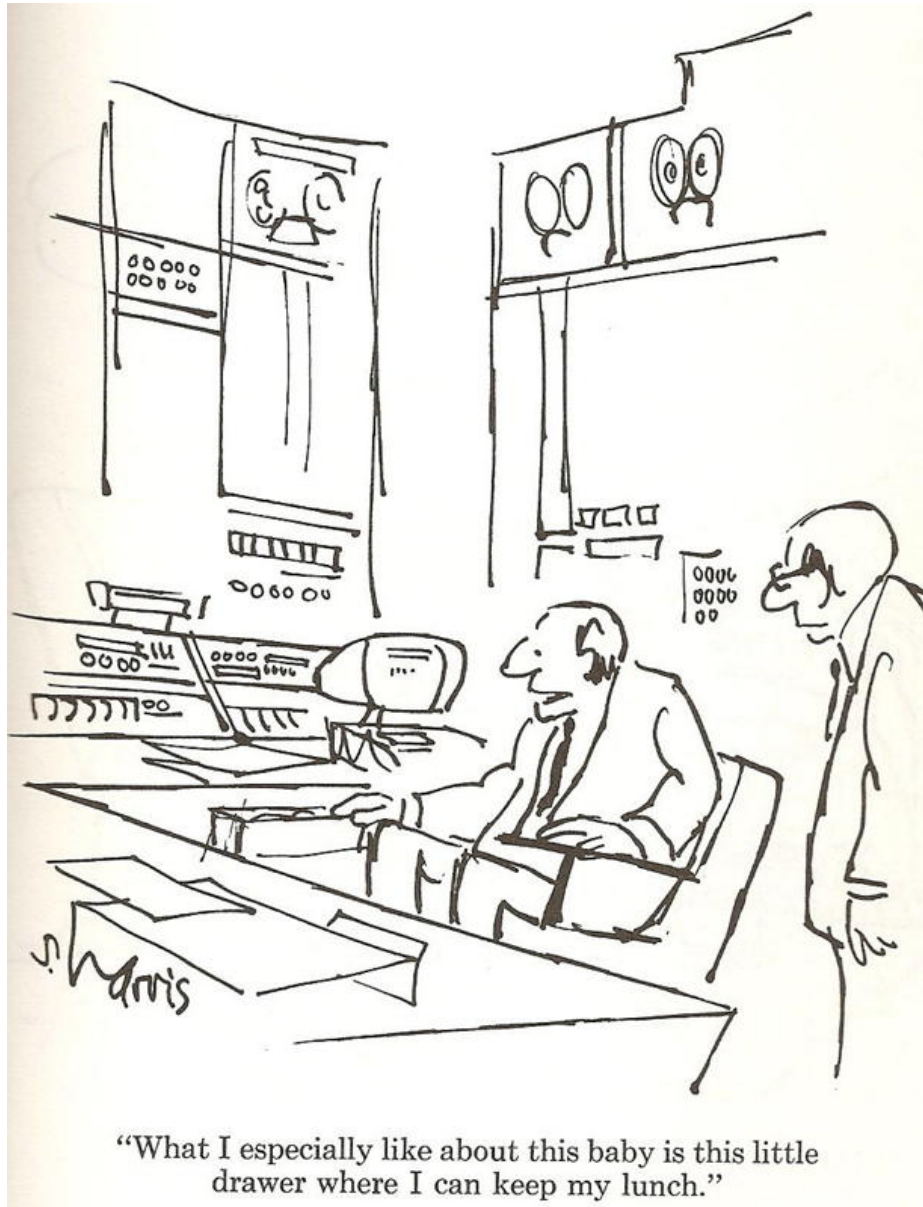
RNA-Seq: *How to Choose?*



Illumina



Ion Torrent



"What I especially like about this baby is this little drawer where I can keep my lunch."



Helicos



Oxford Nanopore

Generating RNA-Seq: *How to Choose?*

Popular choices for RNA-Seq today



Illumina



454



SOLiD



Helicos



Ion Torrent



Pacific Biosciences



Oxford Nanopore

Generating RNA-Seq: *How to Choose?*

Popular choices for RNA-Seq today

[Current RNA-Seq
workhorse]



Illumina



[Full-length single
molecule sequencing]



Pacific Biosciences

[Newly emerging
technology for full-length
single molecule sequencing]



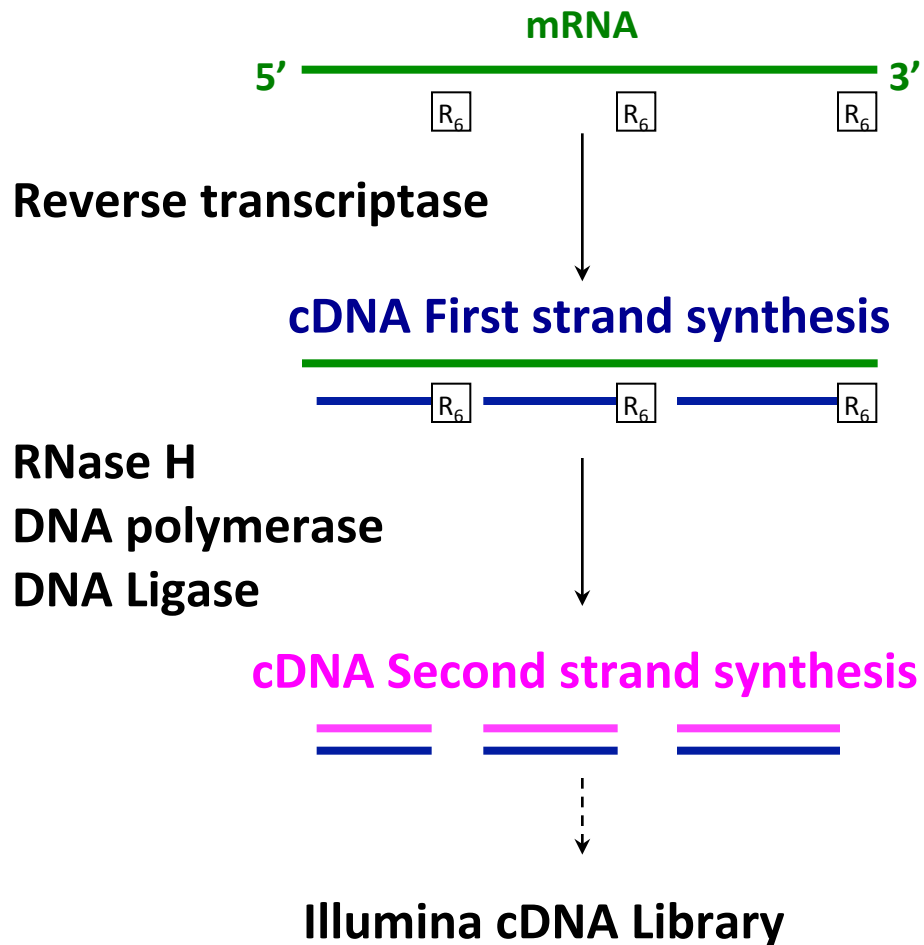
Oxford Nanopore



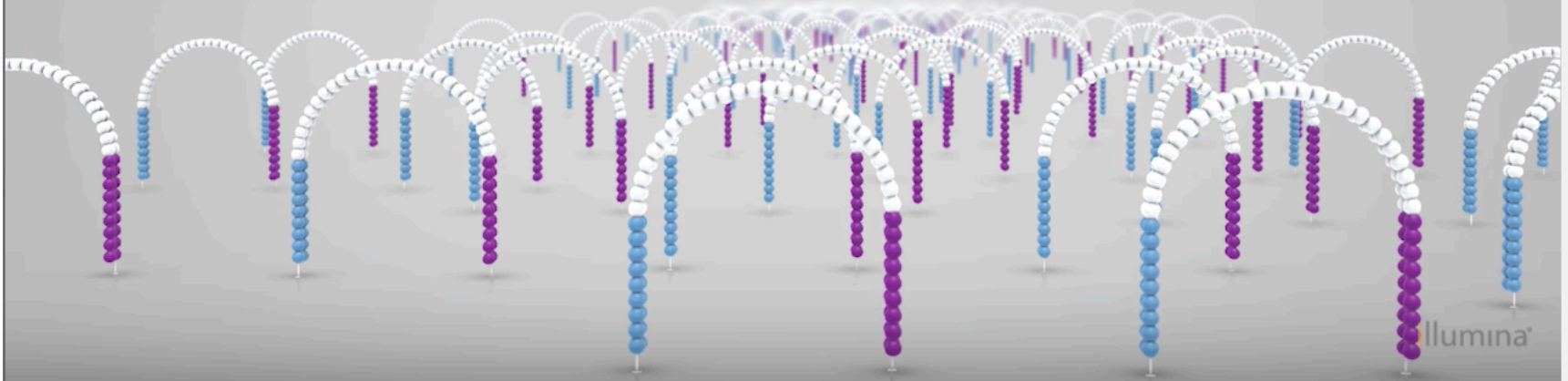
Ion Torrent

RNA-Seq: How do we make cDNA?

Prime with Random Hexamers (R6)



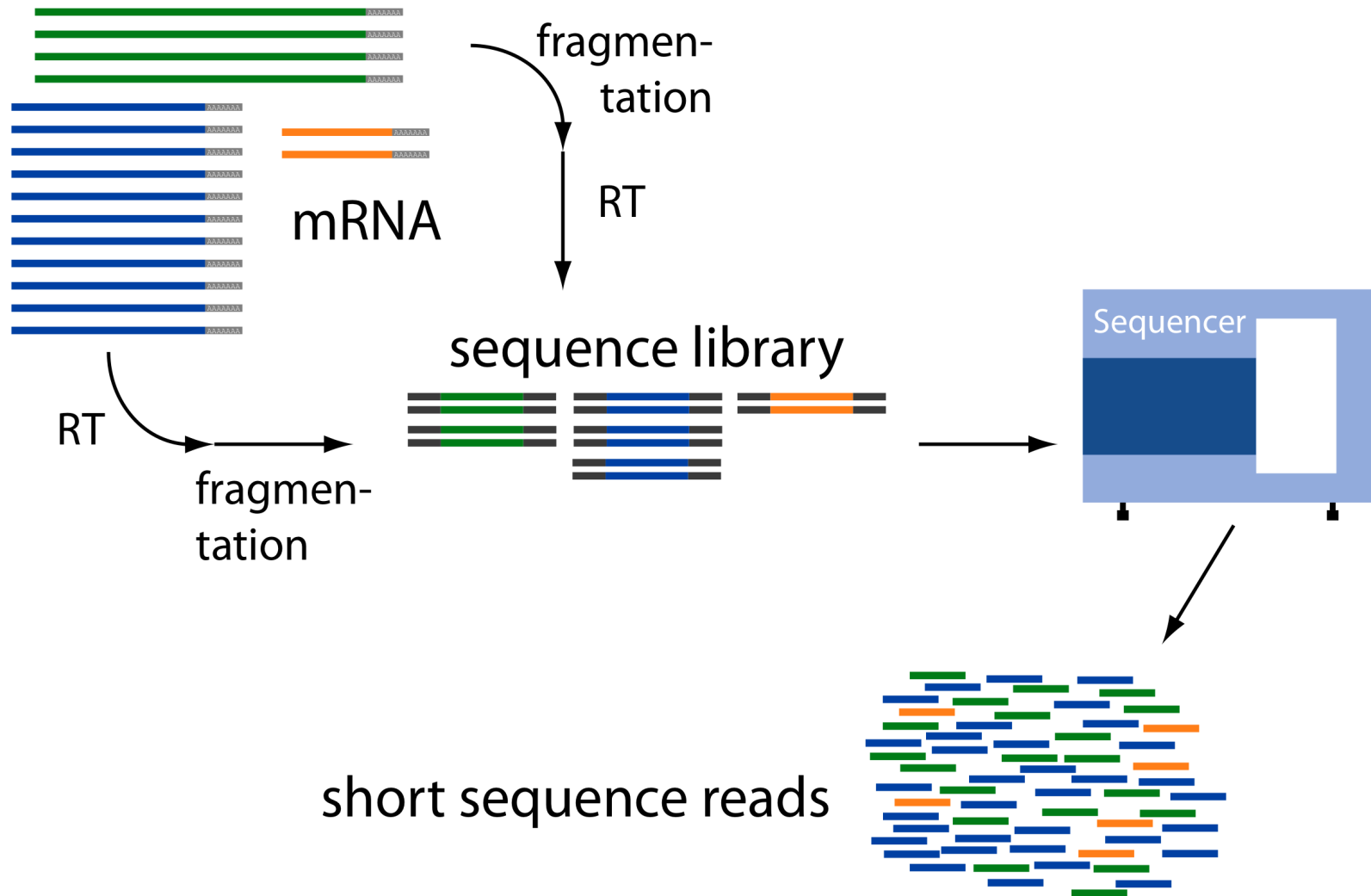
Cluster Generation



2:01 / 5:12



Overview of RNA-Seq



Common Data Formats for RNA-Seq

FASTA format:

```
>61DFRAAXX100204:1:100:10494:3070/1  
AAACAACAGGGCACATTGTCACTCTTGTATTTGAAAAACACTTTCCGGCCAT
```

FASTQ format:

```
@61DFRAAXX100204:1:100:10494:3070/1  
AAACAACAGGGCACATTGTCACTCTTGTATTTGAAAAACACTTTCCGGCCAT  
+  
ACCCCCCCCCCCCCCCCCCCCCCCCCCCCCBC?CCCCCCCC@@@CACCCCCA
```

Read

Quality values

Interpreting Base Quality Values

@61DFRAAXX100204:1:100:10494:3070/1	
AAACAACAGGGCACATTGTCACTCTTGTATTTGAAAAACACTTTCCGGCCAT	Read
+	
ACCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCBB?CCCCCCCC@@CACCCCCA	Quality values

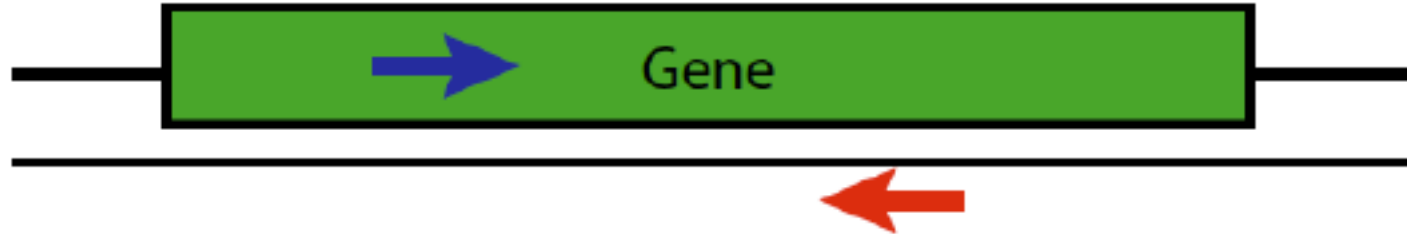
AsciiEncodedQual ('B') = **63**

$$\text{Phred_Quality_Value} = \text{AsciiEncodedQual}('B') - 33 = 30$$

$$\text{Phred_Quality_Value} = -10 * \log_{10}(\text{Pwrong}('T'))$$

$$\text{Pwrong}('T') = 10^{(30/-10)} = 10^{-3} = 0.001$$

Paired-end Sequences

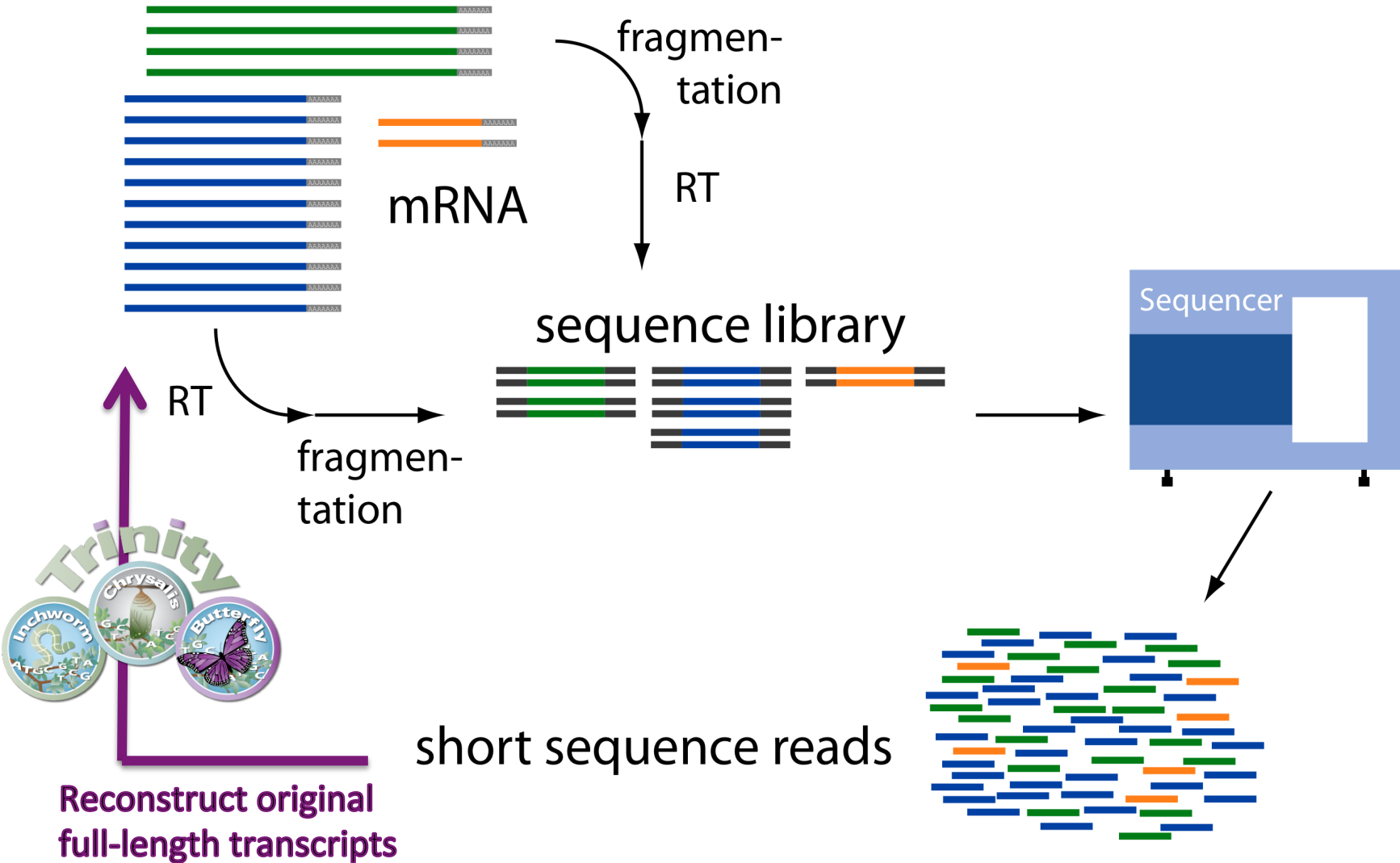


Two FastQ files, read name indicates left (/1) or right (/2) read of paired-end

```
@61DFRAAXX100204:1:100:10494:3070/1  
AAACAACAGGGCACATTGTCACTCTTGTATTTGAAAAACACTTTCCGGCCAT  
+  
ACCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCBC?CCCCCCCC@@@CACCCCCA
```

```
@61DFRAAXX100204:1:100:10494:3070/2  
CTCAAATGGTTAATTCTCAGGCTGCAAATATTCGTTTCAGGATGGAAGAACA  
+  
C<CCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCBCCCC
```

Overview of RNA-Seq



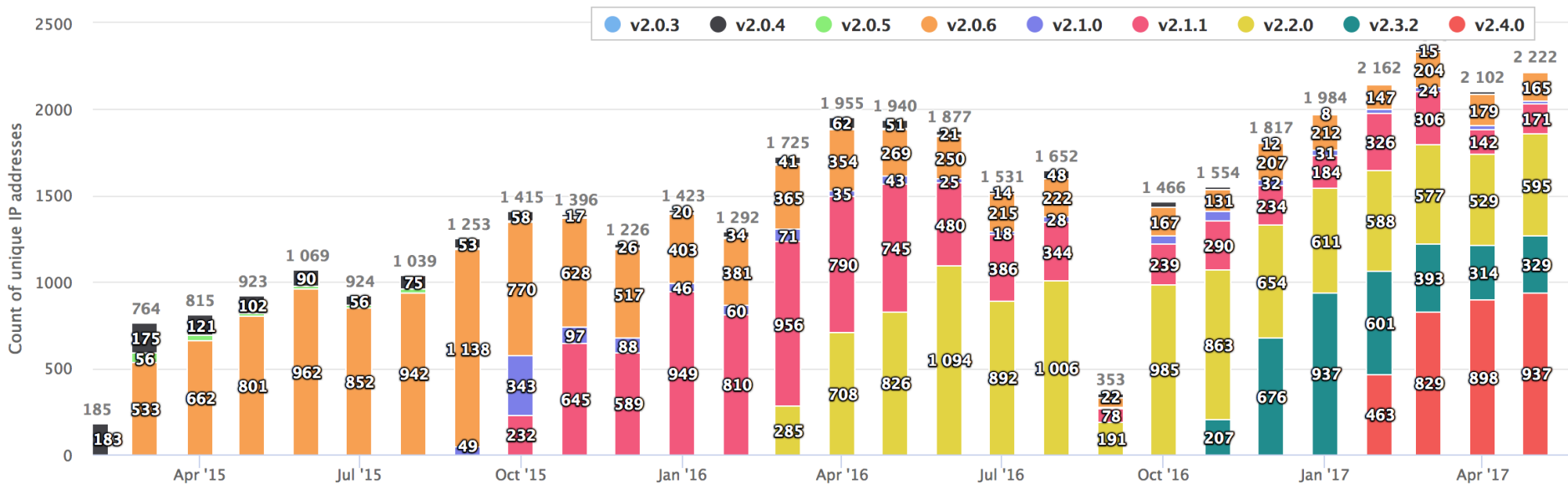
The Ever-Growing Trinity User Community



- ~2k unique users per month
- >4k literature citations (~20% cancer community)
- Open Source software development contributions from the Trinity community.

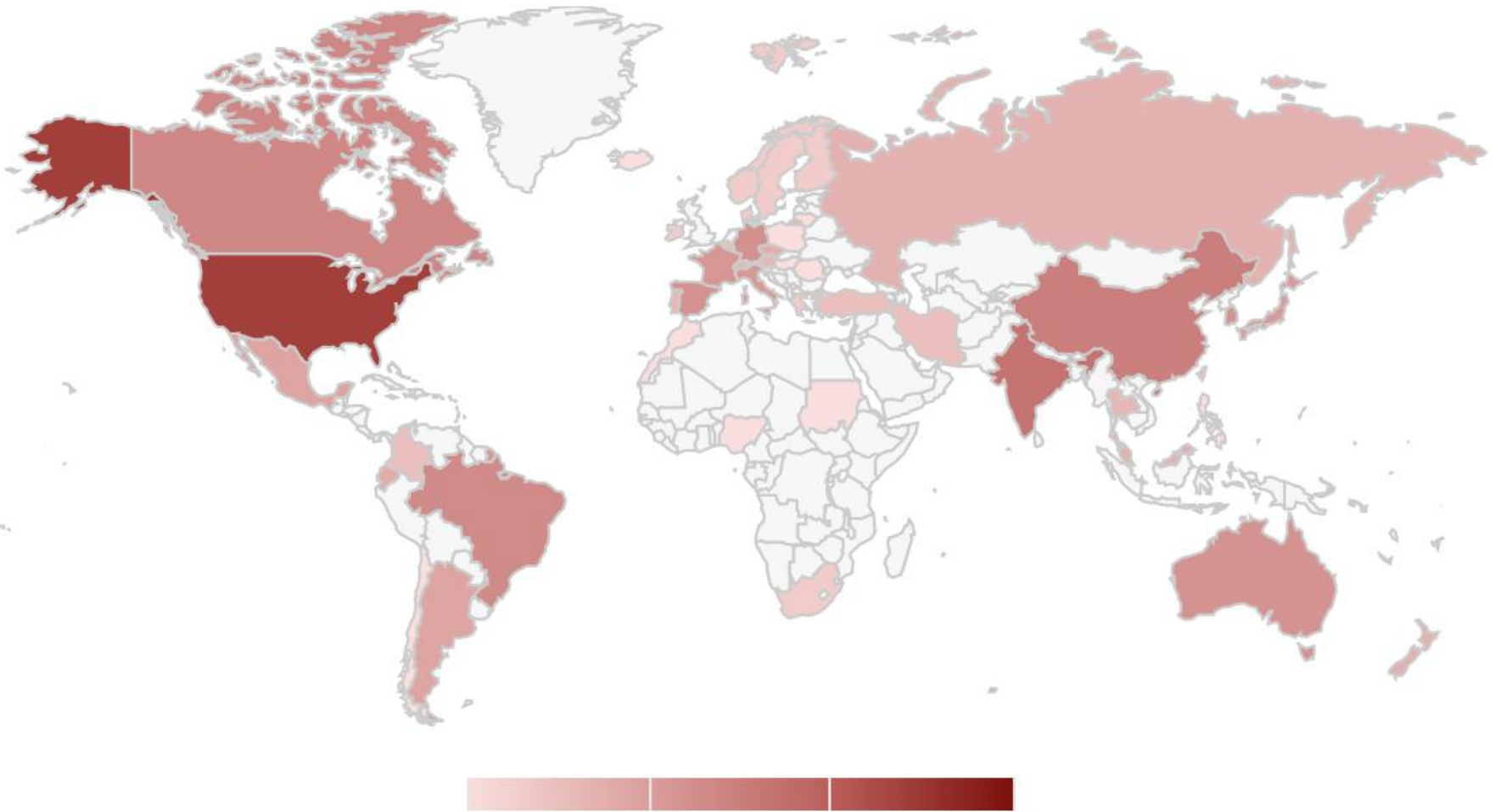


Trinity Usage Tracked by Unique IP Address



Trinity Galaxy Users 2017

Use at 486 institutions in 51 countries



User support and training:

1 10 100 1k

- Google group for community interaction and support.
- Extensive documentation, user guides, tutorials and protocols
- Demo and training videos
- On-site training workshops

RNA-Seq De novo Assembly Using Trinity

Pages 27



Quick Guide for the Impatient

Trinity assembles transcript sequences from Illumina RNA-Seq data.

Download Trinity [here](#).

Build Trinity by typing 'make' in the base installation directory.

Assemble RNA-Seq data like so:

```
Trinity --seqType fq --left reads_1.fq --right reads_2.fq --CPU 6 --max_memory 20G
```

Find assembled transcripts as: 'trinity_out_dir/Trinity.fasta'

Use the documentation links in the right-sidebar to navigate this documentation, and contact our [Google group for technical support](#).

- [Trinity Wiki Home](#)
- [Installing Trinity](#)
 - [Trinity Computing Requirements](#)
 - [Accessing Trinity on Publicly Available Compute Resources](#)
 - [Run Trinity using Docker](#)
- [Running Trinity](#)
 - [Genome Guided Trinity Transcriptome Assembly](#)
 - [Gene Structure Annotation of Genomes](#)
- [Trinity process and resource monitoring](#)
 - [Monitoring Progress During a Trinity Run](#)
 - [Examining Resource Usage at the End of a Trinity Run](#)
- [Output of Trinity Assembly](#)
- [Assembly Quality Assessment](#)
 - [Counting Full-length Transcripts](#)
 - [RNA-Seq Read Representation](#)
 - [Contig Nx and ExN50 stats](#)
 - [Examine strand-specificity of reads](#)
- [Downstream Analyses](#)

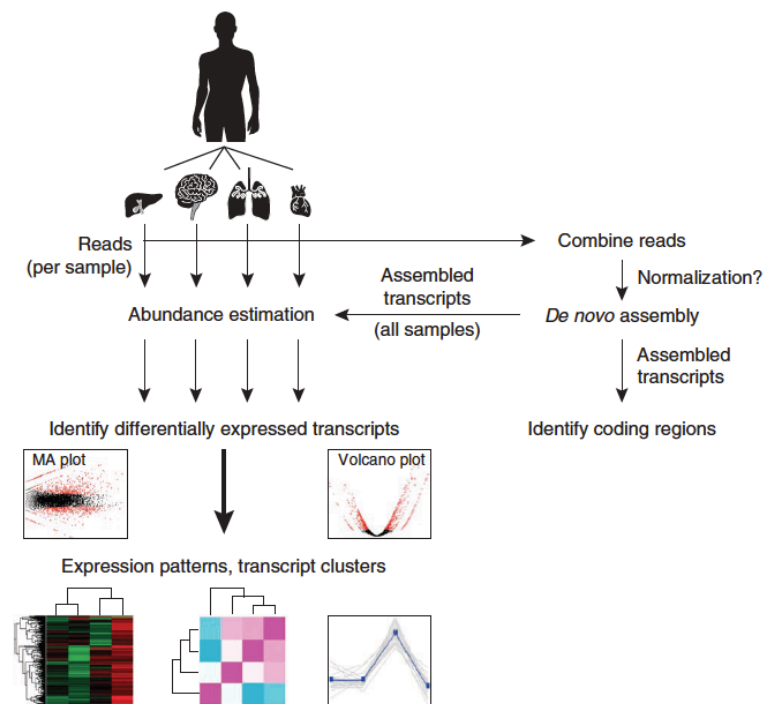
De novo transcript sequence reconstruction from RNA-seq using the Trinity platform for reference generation and analysis

Brian J Haas, Alexie Papanicolaou, Moran Yassour, Manfred Grabherr, Philip D Blood, Joshua Bowden, Matthew Brian Couger, David Eccles, Bo Li, Matthias Lieber, Matthew D MacManes, Michael Ott, Joshua Orvis, Nathalie Pochet, Francesco Strozzi, Nathan Weeks, Rick Westerman, Thomas William, Colin N Dewey, Robert Henschel, Richard D LeDuc, Nir Friedman & Aviv Regev

[Affiliations](#) | [Contributions](#) | [Corresponding authors](#)

Nature Protocols **8**, 1494–1512 (2013) | doi:10.1038/nprot.2013.084

Published online 11 July 2013



Framework for De novo Transcriptome Assembly and Analysis

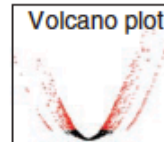
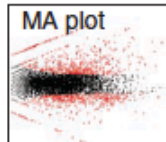


Reads
(per sample)

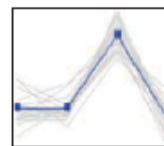
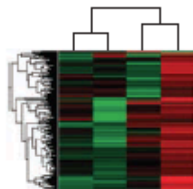
Abundance estimation

Bowtie & RSEM

Identify differentially expressed transcripts



Expression patterns, transcript clusters



Assembled
transcripts
(all samples)



Combine reads

Normalization?

De novo assembly

Assembled
transcripts

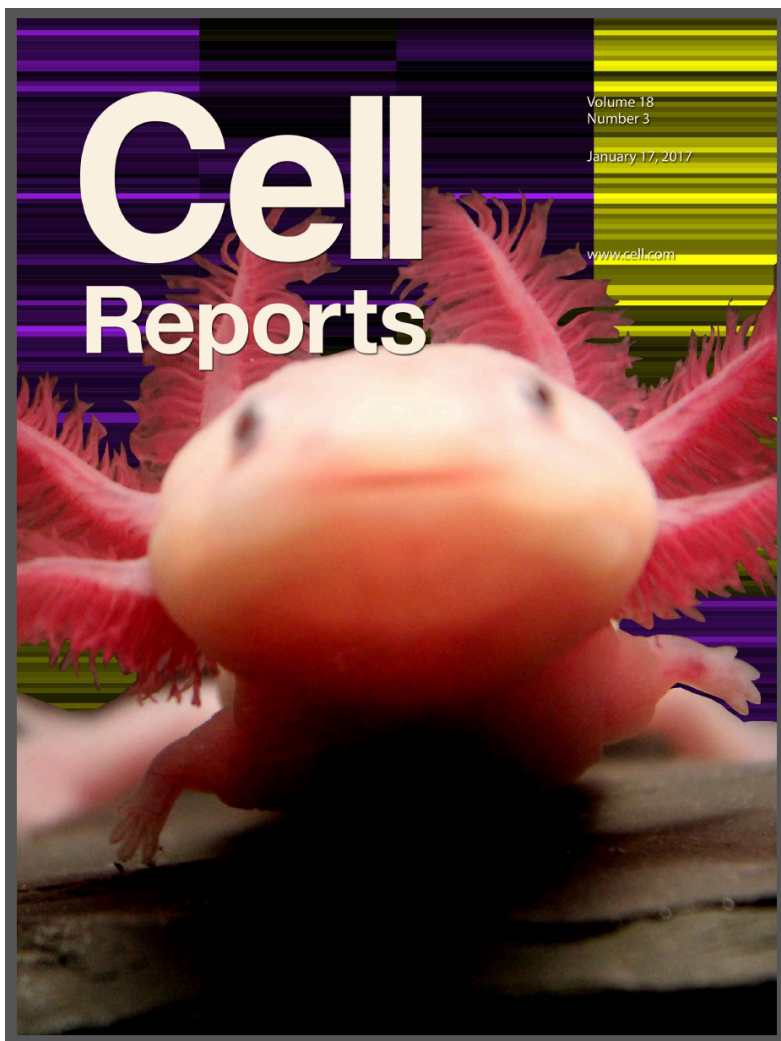
Identify coding regions

1.3 Billion
Total Reads

86 Million
Normalized Reads

EdgeR,
Bioconductor,
& Trinity

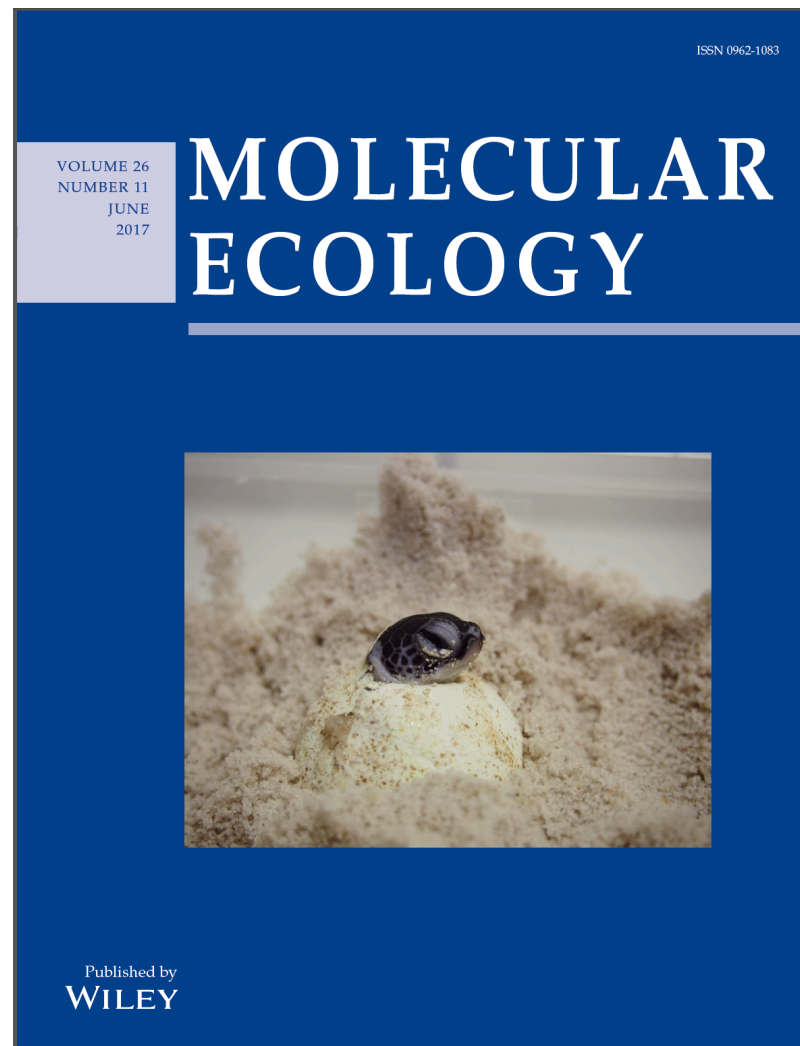
Example Applications of the Trinity RNA-Seq Protocol



Resource

A Tissue-Mapped Axolotl De Novo Transcriptome Enables Identification of Limb Regeneration Factors


Donald M. Bryant^{1,6}, Kimberly Johnson^{1,6}, Tia DiTommaso¹, Timothy Tickle², Matthew Brian Couger³, Duygu Payzin-Dogru¹, Tae J. Lee¹, Nicholas D. Leigh¹, Tzu-Hsing Kuo¹, Francis G. Davis¹, Joel Bateman¹, Sevara Bryant¹, Anna R. Guzikowski¹, Stephanie L. Tsai⁴, Steven Coyne¹, William W. Ye¹, Robert M. Freeman Jr.⁵, Leonid Peshkin⁵, Clifford J. Tabin⁴, Aviv Regev², Brian J. Haas²,  , Jessica L. Whited^{1,7}.



Published by
WILEY

Original Article

Loggerhead sea turtle embryos (*Caretta caretta*) regulate expression of stress response and developmental genes when exposed to a biologically realistic heat stress

Blair P. Bentley , Brian J. Haas, Jamie N. Tedeschi, Oliver Berry

Got RNA-Seq?



Run Trinity