

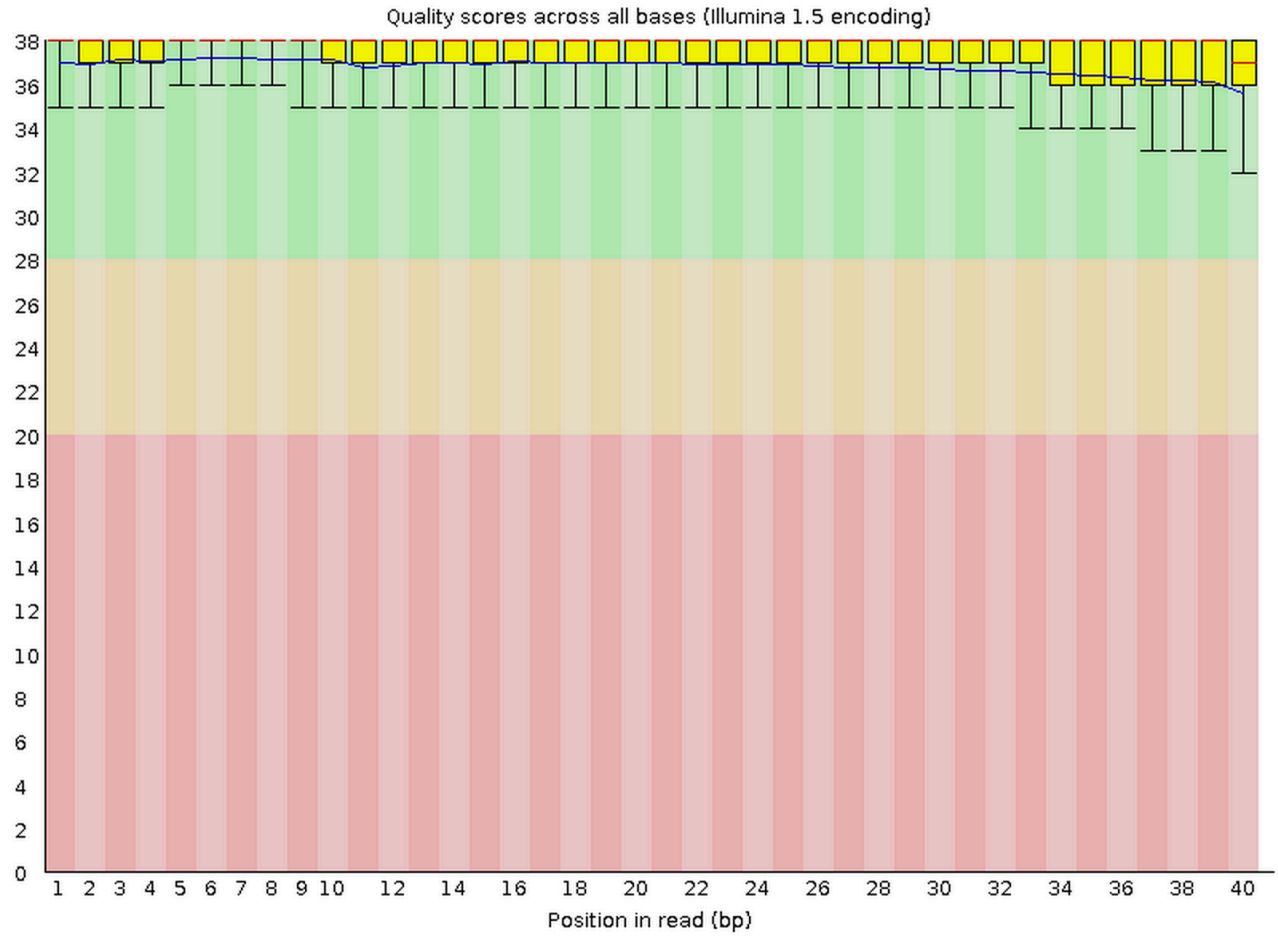
FastQC Report

Wed 25 Mar 2015
good_sequence_short.txt

Summary

- ✔ [Basic Statistics](#)
- ✔ [Per base sequence quality](#)
- ✔ [Per tile sequence quality](#)
- ✔ [Per sequence quality scores](#)
- ! [Per base sequence content](#)
- ✔ [Per sequence GC content](#)
- ✔ [Per base N content](#)
- ✔ [Sequence Length Distribution](#)
- ✔ [Sequence Duplication Levels](#)
- ✔ [Overrepresented sequences](#)
- ✔ [Adapter Content](#)
- ! [Kmer Content](#)

✔ Per base sequence quality

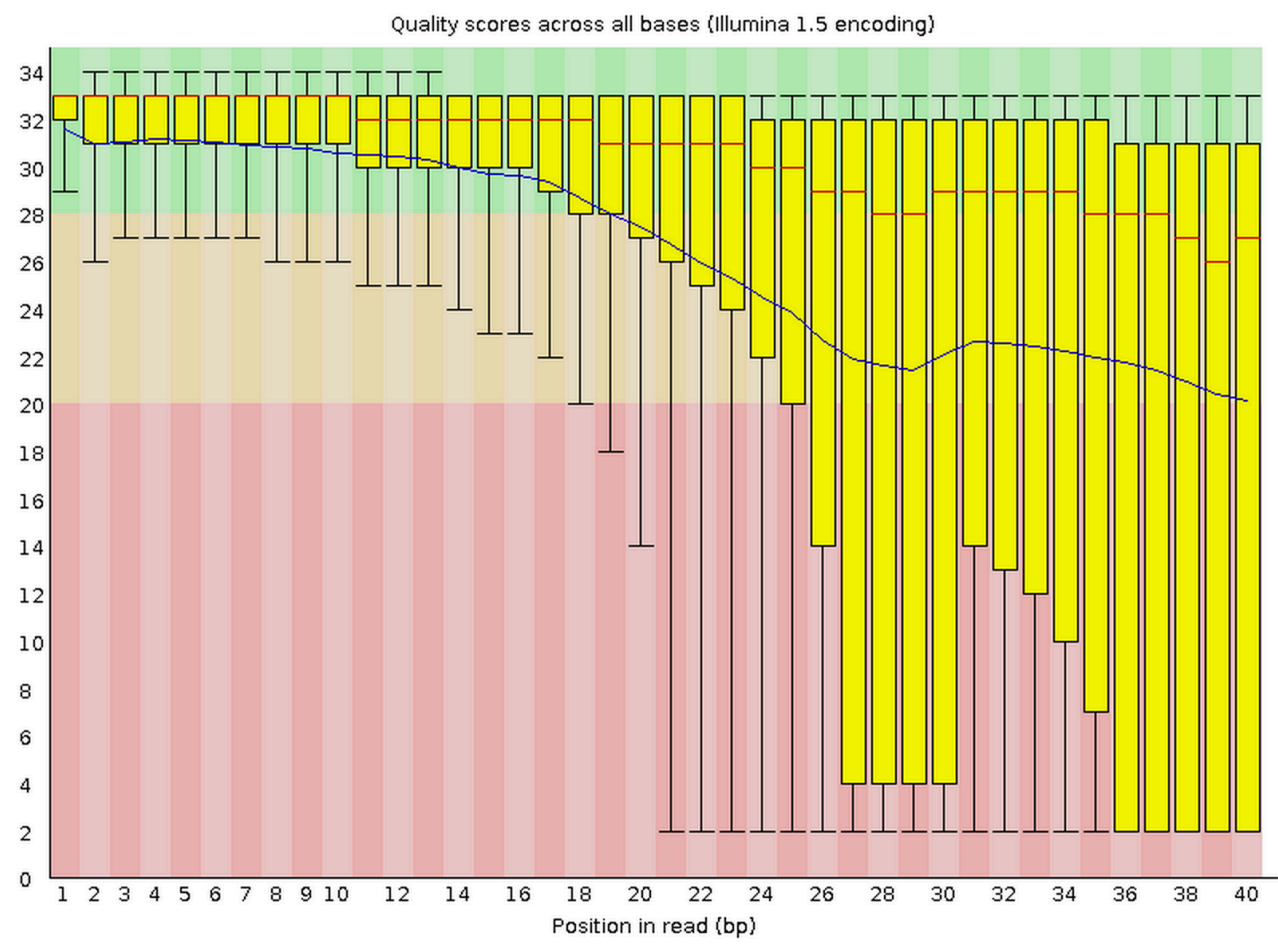


FastQC Report

Summary

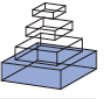
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✘ Per base sequence quality



What to do?

- Trim the reads?
- Start over – try sequencing it again?



On the optimal trimming of high-throughput mRNA sequence data

Matthew D. MacManes^{1,2*}

¹ Department of Molecular, Cellular and Biomedical Sciences, University of New Hampshire, Durham, NH, USA

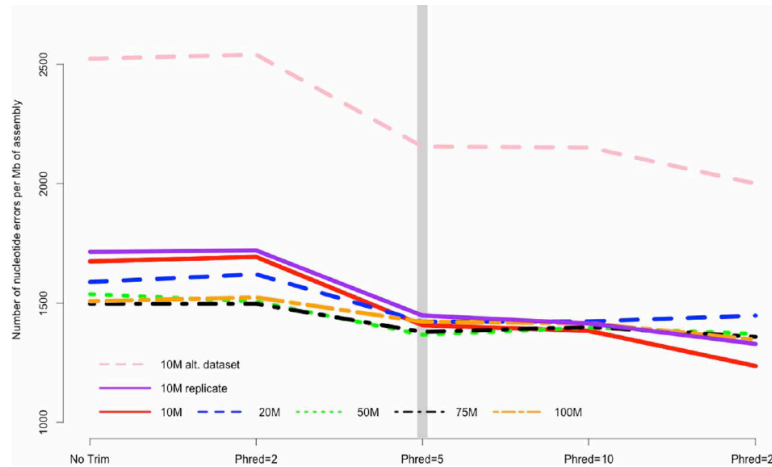
² Hubbard Center for Genome Studies, Durham, NH, USA

“... researchers interested in assembling transcriptomes de novo should elect for a much gentler quality trimming, or no trimming at all.”

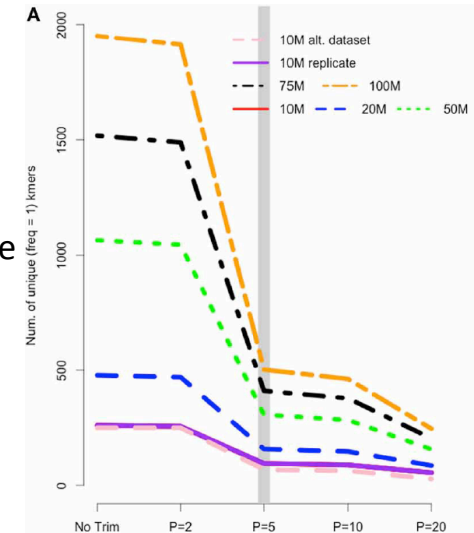
“... trimming at PHRED=2 or PHRED=5 optimizes assembly quality.”

Aggressive Trimming may be harmful, whereas light trimming could be beneficial

Fewer errors in the assembly



Fewer unique kmers

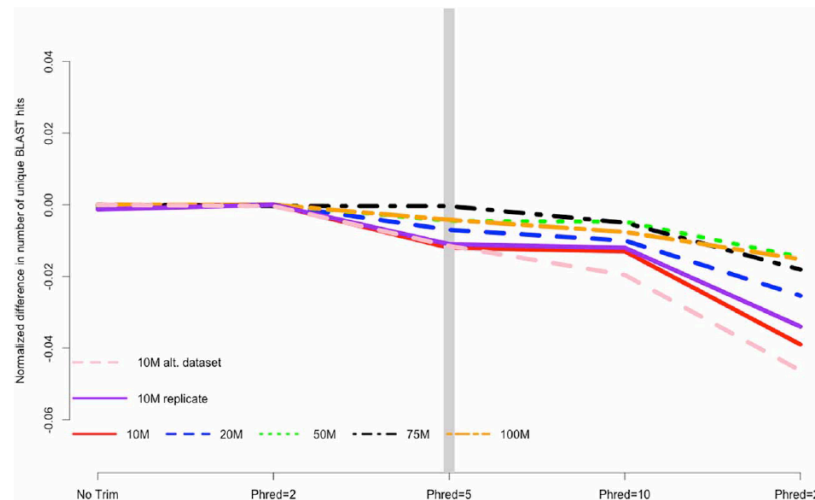


Nucleotide errors / Mb

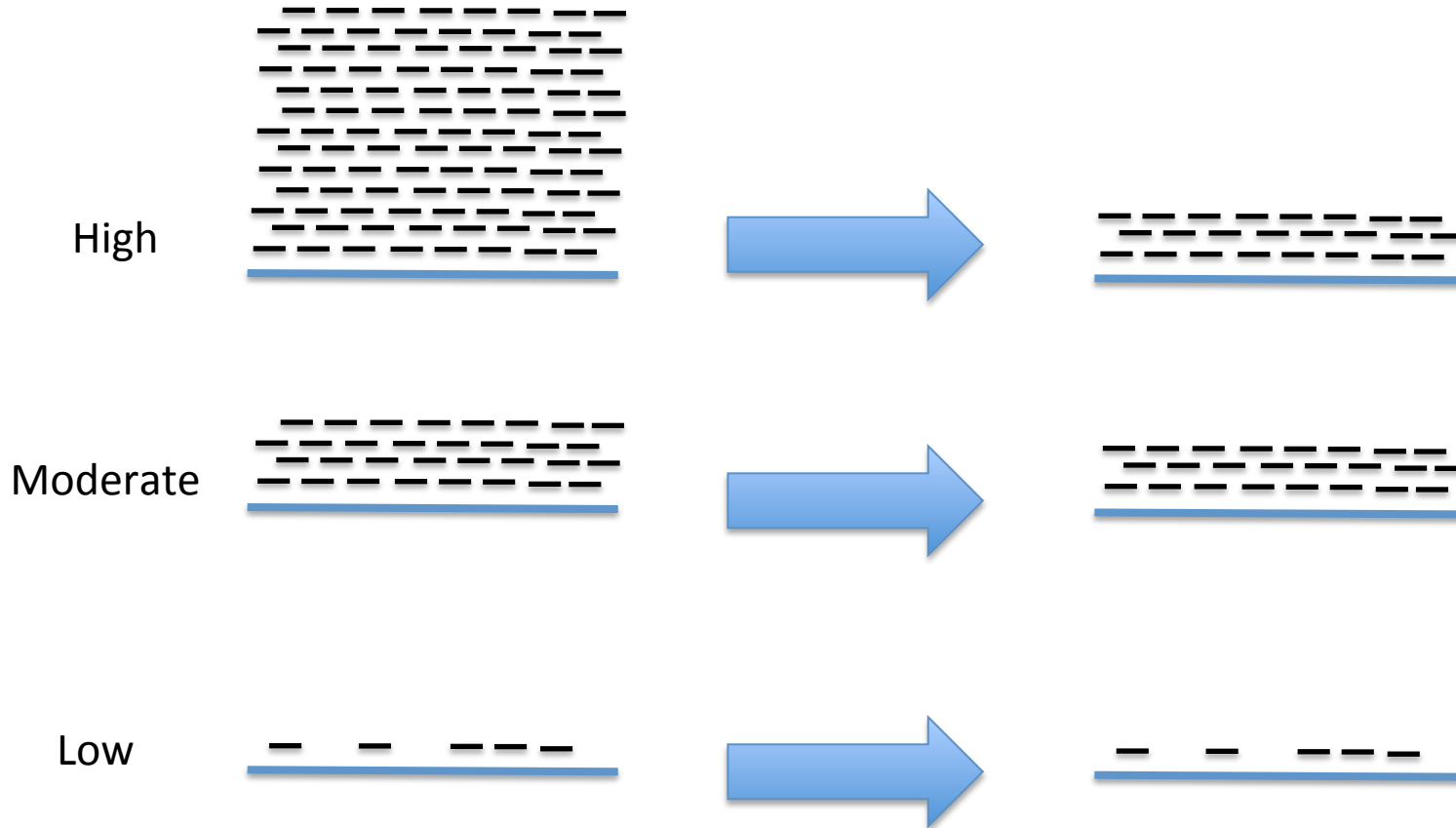
unique kmers

Light trimming doesn't reduce number of blast matches w/ higher sequencing depths.

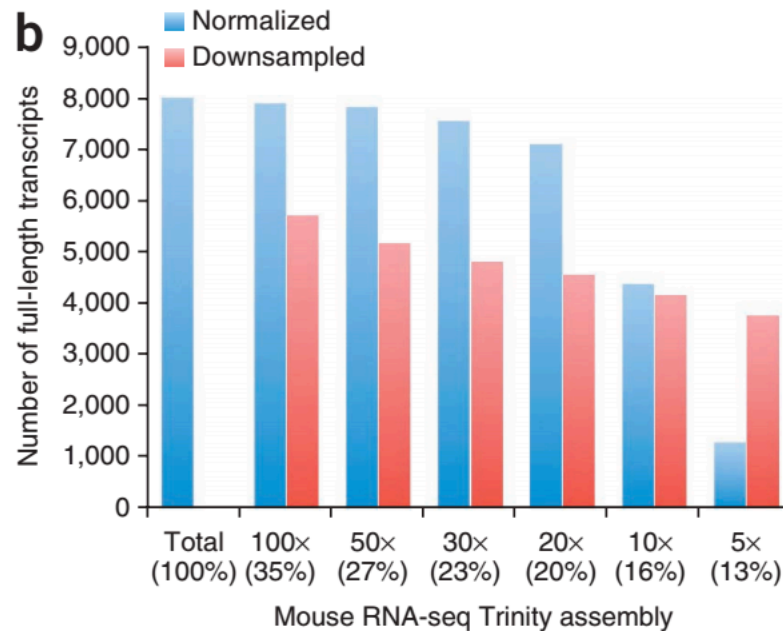
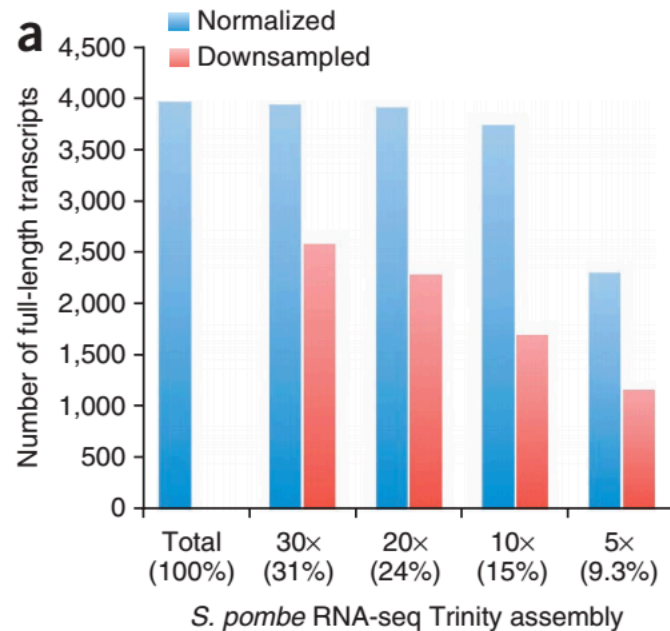
Normalized # of blast matches



In silico normalization of reads



Impact of Normalization on *De novo* Full-length Transcript Reconstruction



Largely retain full-length reconstruction, but use less RAM and assemble much faster.

Quality Trimming and Normalization via Trinity

- Quality Trimming using Trimmomatic:
 - Trinity --trimmomatic
- Normalization of reads:
 - Trinity --normalize_reads *(now on by default!)*
- You can do both in a single Trinity assembly run:
 - Trinity --trimmomatic --normalize_reads

Fastqc, trimming, and normalization practical